



# HHS Public Access

Author manuscript

*Am J Obstet Gynecol.* Author manuscript; available in PMC 2019 April 01.

Published in final edited form as:

*Am J Obstet Gynecol.* 2018 April ; 218(4): 379–389. doi:10.1016/j.ajog.2017.08.010.

## Diet and Fertility: A Review

**Audrey J. GASKINS, Sc.D.<sup>1,2</sup>** and **Jorge E. CHAVARRO, M.D., Sc.D.<sup>1,2,3</sup>**

<sup>1</sup>Department of Nutrition, Harvard T.H. Chan School of Public Health, Boston, MA

<sup>2</sup>Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA

<sup>3</sup>Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, MA

### Abstract

The literature on the relation between diet and human fertility has greatly expanded over the last decade resulting in the identification of a few clear patterns. Intake of supplemental folic acid, particularly at doses higher than those recommended for the prevention of neural tube defects, has been consistently related to lower frequency of infertility, lower risk of pregnancy loss and greater success in infertility treatment. On the other hand, and despite promising evidence from animal models, vitamin D does not appear to exert an important role in human fertility in the absence of deficiency. Antioxidant supplementation does not appear to offer any benefits to women undergoing infertility treatment, but it appears to be beneficial when it is the male partner who is supplemented. However, the available evidence does not allow discerning which specific antioxidants, nor at which doses, are responsible for this benefit. Long chain omega 3 fatty acids appear to improve female fertility although it remains unclear to what extent contamination of shared food sources, such as fish with high levels of environmental toxicants, can dampen this benefit. Last, adherence to healthy diets favoring seafood, poultry, whole grains, fruits and vegetables, are related to better fertility in women and better semen quality in men. The cumulative evidence has also piled against popular hypotheses. Dairy and soy, once proposed as reproductive toxicants, have not been consistently related to poor fertility. In fact, soy and soy supplements appear to exert a beneficial effect among women undergoing infertility treatment. Similarly, as data from large, high-quality studies continues to accumulate, the evidence of a potentially deleterious effect of moderate alcohol and caffeine intake on the ability to become pregnant seems less solid than it once did. While a complete picture of the role of nutrition on fertility is far from complete, much progress has been made. The most salient gaps in the current evidence include jointly considering female and male diets, and testing the most consistent findings in randomized trials.

---

Corresponding Author: Jorge E. Chavarro, M.D., Sc.D., 655 Huntington Avenue, Building II 3rd Floor, Boston, MA 02115, Phone: (617) 432-4584 jchavarr@hsph.harvard.edu.

**Conflicts of Interest:** The authors report no conflicts of interest.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

## Keywords

nutrition; diet; fertility; fecundity; spontaneous abortion; miscarriage; pregnancy loss; in vitro fertilization; assisted reproduction; reproductive health

---

## Introduction

Identifying modifiable lifestyle factors, such as diet, that influence human fertility is of major clinical and public health significance. Infertility, the failure to achieve a clinical pregnancy after 12 months or more of regular unprotected sexual intercourse, affects 15–25% of couples in western countries.<sup>1, 2</sup> Impaired fecundity, which encompasses infertility and difficulty carrying a pregnancy to term, is estimated to affect twice as many couples.<sup>3</sup> Medical treatment for impaired fecundity is also on the rise. The use of assisted reproductive technologies (ART) in the United States has steadily increased from approximately 60,000 cycles in 1995<sup>4</sup> to 209,000 cycles in 2015,<sup>5</sup> although improvements in live birth rates per initiated cycle over the last decade have been small in comparison. Comparable data for intrauterine insemination and ovulation induction procedures are lacking but given that these procedures are even more common, it's estimated they account for 2–6 times more births than ART in the United States.<sup>6</sup>

The high prevalence of impaired fecundity combined with the high financial costs of and limited geographic access to infertility treatment motivate the need to identify modifiable predictors of couple fertility.<sup>7, 8</sup> While there is a growing acceptance that nutrition may be related to reproductive performance in both men and women,<sup>9</sup> there is still no official guidance for reproductive-aged couples. The purpose of this review is to summarize the epidemiologic literature on nutrition and fertility and offer practical dietary recommendations based on the best available evidence. Highlights and gaps in the literature are summarized in Table 1 for female diet and Table 2 for male diet.

## Micronutrients

### Antioxidants

A 2013 Cochrane review of randomized controlled trials (RCTs) of antioxidant supplementation during the course of infertility treatment concluded that the current evidence does not show benefits of antioxidant supplementation for increasing pregnancy or live birth rates.<sup>10</sup> The authors pointed out many deficiencies of the available evidence including high risk of bias, incomplete reporting, and high variability of the interventions tested in the trials.<sup>10</sup> For example, the trials included in the meta-analysis testing the effect of “antioxidants” against placebo included interventions as dissimilar as multiple micronutrient blends (including proprietary blends with undisclosed ingredients), pentoxifyline, *N*-acetyl-cysteine, melatonin, L-arginine, vitamin E, myo-inositol, vitamin C, vitamin D+calcium and omega-3 polyunsaturated fatty acids, many of which are not even technically antioxidants. Furthermore, no two trials included in the meta-analysis tested the same intervention (i.e. same compound at same dose against same comparator) making it nearly impossible to draw strong conclusions from this systematic review other than the

need of more high quality trials large enough to test effects on clinically relevant outcomes such as live birth rates.

## B Vitamins

More promising nutrients in the context of beneficial effects on fertility might possibly be folate (or folic acid) and vitamin B12. While the impact of folate deficiency and defects in folate and homocysteine metabolism on neural tube defects (NTDs) are established,<sup>11</sup> the evidence on the effects of folate on fertility is less clear. One of the first studies supporting a link between folate and fertility was the Hungarian NTDs RCT which showed that of the women randomized to the pre-conception multivitamin supplement (containing 800 µg of folic acid) 71.3% conceived compared 67.9% of the women randomized to the placebo-like trace element supplement during a 14-month follow-up period.<sup>12</sup> Similarly, in a small RCT, of the subfertile women who took a multivitamin (containing 400 µg of folic acid) for 3 months, 26% had a pregnancy compared to 10% of women in the placebo group.<sup>13</sup> Among women participating in the Nurses' Health Study II (NHS-II) cohort, women who consumed 6 multivitamin tablets per week had a 41% (95% CI 25, 54%) lower risk of ovulatory infertility compared to non-consumers with folic acid appearing to explain most of this association.<sup>14</sup> Moreover, it was estimated that 20% (95% CI 11, 28%) of the ovulatory infertility cases could be avoided if women consumed 3 or more multivitamins per week. Consistent with this finding, folate intake was related to a lower frequency of sporadic anovulation in a prospective cohort of young healthy women (adjusted odds ratio=0.36 [95% CI 0.14, 0.92] comparing women in highest to lowest tertile of folic acid).<sup>15</sup> Most recently, folic acid supplement use was also associated with shorter time to pregnancy among a large cohort of Danish pregnancy planners (adjusted fecundability ratio=1.15 [95% CI 1.06, 1.25]).<sup>16</sup>

Studies from infertility cohorts also suggest that folate could have beneficial effects on fertility. For instance, carriers of the T allele in position 677 of the MTHFR gene (which leads to lower enzyme activity) had decreased ovarian responsiveness to follicle-stimulating hormone, fewer oocytes retrieved,<sup>17</sup> and granulosa cells that produced less estradiol (basal and stimulated) compared to wild type allele carriers.<sup>18</sup> A Polish *in vitro* fertilization (IVF) cohort study also found that women who received a folic acid supplement prior to treatment had better quality oocytes and a higher degree of mature oocytes compared to women who did not receive folic acid.<sup>19</sup> Similarly, among a cohort of US women undergoing IVF, with nearly universal compliance to preconception folic acid supplement use guidelines and no evidence of folate or B12 deficiency, the probability of live birth was 20% (8, 31%) higher among women consuming >800 µg/day of supplemental compared to women consuming <400 µg/day. Similarly, in this same cohort, women in the highest quartile of serum folate and vitamin B12 levels had 1.62 (95% CI 0.99, 2.65) and 2.04 (95% CI 1.14, 3.62) times the probability of live birth compared to women in the lowest quartiles.<sup>20, 21</sup> While three other cohort studies of folate and clinical outcomes of IVF from European populations did not show this benefit,<sup>22-24</sup> they excluded women failing prior to embryo transfer, which could have systematically biased their findings towards the null if folate does indeed prevent early failures prior to embryo transfer, as suggested by findings from the US cohort<sup>20</sup> and Dutch studies relating markers of folate and B12 status with greater day 3 embryo quality.<sup>25,26</sup>

Nevertheless, there has been plenty of controversy surrounding the evidence on folate and pregnancy maintenance. Only a couple years after the Hungarian RCT findings were published on the beneficial effects of a multivitamin supplement on likelihood of conception, the full report on all pregnancy outcomes was released which suggested that pre-conception folic acid use increased the risk of fetal death (RR=1.16 [95% CI 1.01, 1.30]).<sup>27</sup> This was further supported by a re-analysis of the UK Medical Research Council study (RR=1.15, p-value=0.18) and an observational cohort study in California (RR=1.14 [95% CI 0.96, 1.35]) which also found similar increased risks (albeit not statistically significant).<sup>28, 29</sup> These results were later challenged due to methodological errors<sup>30, 31</sup> and a subsequent trial from India did not replicate these findings (RR=0.44).<sup>32</sup> Thus, the most recent Cochrane review concluded that there was no evidence across three randomized and quasi-randomized trials of any difference in the risk of total fetal loss (RR=1.00 [95% CI 0.75, 1.34]), early or late miscarriage (RR=0.99 [95% CI 0.72, 1.38]), or stillbirth (RR=1.03 [95% CI 0.51, 2.09]) comparing women supplemented with folic acid compared to none.<sup>33</sup> Results from observational folic-acid intervention studies in China (RR= 0.97 [95% CI 0.84, 1.12])<sup>34</sup> and Brazil (RR=0.80, p-value=0.49)<sup>35</sup> have also since provided strong evidence that periconceptional folic acid use did not increase miscarriage rates. Moreover, three recent cohort studies have shown that the use of folic acid prior to or during early pregnancy is, in fact, associated with a reduced risk of miscarriage (aOR=0.43 [95% CI 0.30, 0.60] and aOR=0.37 [95% CI 0.19, 0.72] for folic acid or vitamin use during pregnancy versus none; aRR=0.80 [95% CI 0.71, 0.90] for >730 µg/day of supplemental folate vs none),<sup>36-38</sup> and an additional prospective cohort found that increased adherence to preconception multivitamin supplements (one of the main sources of folic acid in the US population) was also related to a lower risk of miscarriage (aHR=0.45 [95% CI 0.25, 0.80]).<sup>39</sup>

The vast literature on folate and fertility endpoints suggests that higher intake of preconception supplemental folate may increase a woman's chances of becoming pregnant and, possibly, to carry a pregnancy to term. Interestingly, in several of these studies, beneficial effects of folate on fertility and fecundity were observed at levels well above those that are currently recommended for the prevention of NTDs.

## Vitamin D

Over the last decade, the potential effects of vitamin D on fertility have been of great research interest as *in vitro* studies found that the vitamin D receptor is expressed in the ovary,<sup>40, 41</sup> the endometrium,<sup>40</sup> and the placenta.<sup>42</sup> Animal studies have also pointed to a possible role of vitamin D in fertility as female rodents fed a vitamin D deficient diets and female rodents with knockouts for *VDR* and *1 $\alpha$ -hydroxylase* (which catalyses the hydroxylation of 25(OH)D into the biologically active 1,25(OH)<sub>2</sub>D) were shown to have reduced fertility<sup>43, 44</sup> as a result of uterine hypoplasia, impaired follicular development and anovulation.<sup>45-47</sup>

Among women trying to get pregnant in the NHS-II cohort, higher intake of vitamin D (as estimated through a food frequency questionnaire) was not associated with risk of ovulatory infertility after multivariable adjustment. Similarly, among a large cohort of women with 1-2 prior pregnancy losses and no history of infertility, there were no associations between

baseline serum vitamin D levels or vitamin D deficiency (<20 ng/mL) and fecundability.<sup>48</sup> A cohort study among 153 Danish pregnancy planners also found no associations between pre-conception plasma 25-hydroxyvitamin D concentrations and chances of conceiving or overall risk of miscarriage; however, women who had a miscarriage after gestational week 10 had lower first trimester plasma vitamin D concentrations compared with those who did not have a miscarriage.<sup>49</sup> These findings should be interpreted with caution though as it was only based on a small number of cases (n=3) and three other studies found no associations between early pregnancy concentrations of serum 25(OH)D and risk of miscarriage.<sup>50–52</sup> Similarly, a recently published meta-analysis found no association between vitamin D insufficiency and risk of spontaneous abortion (RR=1.04 [95% CI 0.95, 1.13]).<sup>53</sup> Finally, one case-control study, compared early pregnancy levels of vitamin D between women who took 12–24 months to get pregnant compared to age-matched women conceiving in less than 1 year and found no associations.<sup>54</sup>

Despite the limited studies on vitamin D and fertility from the general population, there has been explosion in the number of studies over the past 7 years exploring this association among subfertile women undergoing medical treatment. The first study on vitamin D and fertility after IVF reported that pregnancy rates were almost four fold higher in women who were vitamin D sufficient compared to those who were vitamin D deficient.<sup>55</sup> While a handful of subsequent studies have yielded similar, positive findings,<sup>56–59</sup> a similar number of studies have found no associations<sup>60–64</sup> and one study even observed a negative association.<sup>65</sup> A small randomized controlled trial from Iran among women with insufficient serum vitamin D levels (<30 ng/ml) undergoing a cryopreservation cycle found that vitamin D supplementation of 50,000 IU/week, for 6–8 weeks, was not associated with clinical pregnancy rates.<sup>66</sup> Similarly, a small RCT among PCOS women undergoing intrauterine insemination found no significant differences in pregnancy outcomes<sup>67</sup> despite observational evidence that higher serum vitamin D levels might predict greater reproductive success among PCOS women undergoing ovulation induction.<sup>68</sup> Two studies have investigated the association between vitamin D levels and IVF outcomes among egg donor recipients and while one found a significant increase in clinical pregnancy rates with increasing vitamin D levels<sup>69</sup>, suggesting a specific effect of 25(OH)D on endometrial receptivity, the second study could not confirm this association.<sup>70</sup>

At present, little can be conclusively drawn from the results on vitamin D and fertility given the heterogeneity of findings. While vitamin D deficiency might possibly be detrimental to fertility, it is unclear whether higher levels of vitamin D confer additional benefit once sufficiency has been achieved.

## Fatty Acids

*In vitro* studies have shown that fatty acids are important substrates in early reproductive events including oocyte maturation<sup>71</sup> and embryo implantation.<sup>72</sup> Moreover, animal and human studies suggest that polyunsaturated fatty acids (PUFAs) may specifically impact fertility, through effects on oocyte quality and embryo implantation<sup>73, 74</sup> while trans fatty acids may promote greater insulin resistance<sup>75</sup> which could adversely affect ovulatory function.<sup>76</sup>

Results from the NHS-II cohort demonstrated that *trans* fatty acids (TFA) intake was associated with a greater risk of self-reported ovulatory infertility after adjustment for potential confounders (aRR=1.73 [95% CI: 1.09, 2.73] for a 2% increase in energy from TFA);<sup>77</sup> however intakes of saturated fatty acids (SFAs), monounsaturated fatty acids (MUFAs), total PUFAs, omega 3 PUFAs, and omega 6 PUFAS were not associated with ovulatory infertility. Among two prospective time to pregnancy studies, women in the highest quartile of TFA intake had reduced fecundability in the North American cohort (FR=0.86 [95% CI: 0.71, 1.04]) but not the Danish cohort (FR=1.04 [95% CI: 0.86, 1.25]), although intake in Denmark was low.<sup>78</sup> Additionally, in the North America cohort, women in the lowest quartile of omega-3 PUFA intake had lower fecundability than women in the other quartiles (FR=1.19 [95% CI 1.02, 1.39]) while no association was found in Denmark, where low intake was rare.<sup>78</sup> Docosapentaenoic acid (DPA), an omega 3 PUFA that is structurally similar to eicosapentaenoic acid (EPA), was associated with reduced risk of anovulation in a cohort of healthy, regularly menstruating women (aRR=0.42 [95% CI 0.18, 0.95] for tertile 3 vs. tertile 1), with similar inverse trends for the other long-chain omega 3 fatty acids.<sup>79</sup>

Among 46 overweight and obese women undergoing IVF in Australia, intake of PUFAs, specifically omega-6 PUFAs and linoleic acid (LA) and possibly omega-3 PUFA, was higher among women who achieved pregnancy;<sup>80</sup> however there were no differences in fat intake comparing women who did and did not have live births. Two studies from the US and one from Iran have investigated the association between serum fatty acids concentrations and outcomes of IVF. The first US cohort found that women with lower serum  $\alpha$ -linolenic (ALA) had a higher chance of pregnancy while the second found that only an increased LA to ALA ratio was associated with a higher chance of pregnancy.<sup>81</sup> A cohort study among 105 women underdoing intracytoplasmic sperm injection (ICSI) in Iran found that serum levels of EPA were significantly higher in women who achieved pregnancy compared to those who did not.<sup>82</sup>

While synthesizing these results is difficult given the large differences across studies in terms of populations and assessment of fatty acid status, the conclusions overall appear to suggest that higher intake of PUFAs, specifically long chain omega 3 fatty acids, and lower intake of trans fatty acids may be beneficial for enhancing female fertility.

## Dairy

Dairy foods have been suggested as potential reproductive toxicants due to their high content of galactose, which in mice was shown to decrease ovulation and lead to premature ovarian failure,<sup>83, 84</sup> and their potential to contain high amounts of environmental estrogens.<sup>85</sup> In 1994 an ecological study among 31 countries was published showing that the decline in fertility with age is steeper among populations with higher per capita milk consumption.<sup>86</sup> However, a subsequent case-control study found that women consuming three or more glasses of milk per day had a 70% lower risk of infertility than women who did not consume milk.<sup>87</sup> In NHS-II, the largest prospective cohort to date, no relation was found between total intake of dairy foods and risk of ovulatory infertility (aRR=1.12 [95% CI 0.69, 1.82] comparing 4 vs. <1 serving per day) yet this overall null finding was due to the fact that



full-fat dairy foods were associated with lower risk of ovulatory infertility (aRR=0.73 [95% CI 0.52, 1.01] comparing 1 serving per day vs. 1 servings per week) while low-fat dairy foods were associated with higher risk of ovulatory infertility (aRR=1.85 [95% CI 1.24, 2.77] comparing 2 servings per day vs. 1 servings per week).<sup>88</sup> A prospective cohort of women undergoing assisted reproduction in the US found that among women 35 years of age, those in the highest quartile of pre-treatment dairy food intake had a multivariable-adjusted probability of live birth of 55% (95% CI 39, 69%) compared to 23% (95% CI 11, 42%) among women in the lowest quartile.<sup>89</sup> And while this relationship was only present among older women, the association did not differ between full-fat and low-fat dairy foods.<sup>89</sup> Finally, in the most recent study on pre-conception dairy intake and time to pregnancy, associations between dairy intake and fecundity were small and inconsistent between the Danish and American cohorts (pooled FR=1.11 [95% CI: 0.94, 1.31] comparing 18 vs. <7 servings per week).<sup>90</sup> Taken together, given the conflicting findings, no strong conclusions regarding the effect of maternal dairy intake on fertility can be made although the evidence supporting dairy as a potential reproductive toxicant (similar to animal studies) is weak.

## Meats, fish and soy

Intake of protein sources have received attention in the context of fertility mostly due to their potential to contain high levels of environmental contaminants, which could adversely affect reproductive health. While red meats can be good sources of protein and other essential nutrients, they also contain high levels of saturated fat and can serve as a vehicles for exposure to hormonal residues, antibiotics, and polybrominated diphenyl ethers.<sup>91, 92</sup> Similarly, while seafood is recognized good source of long chain omega 3 fatty acids, it can also be a primary route of exposure to organochlorines, dioxins, and mercury.<sup>93</sup> Moreover, while soy-based products are generally healthy alternatives to animal proteins in terms of cardiovascular and metabolic benefits, some have raised concerns regarding the potential adverse reproductive consequences of soy phytoestrogens.<sup>94</sup>

Among women from the NHS-II cohort, one additional serving of meat (red meats, chicken, turkey, processed meats and fish) per day, while holding calories constant, was associated with a 32% (95% CI 8, 62%) increase in the risk of ovulatory infertility.<sup>95</sup> Similarly, among women from an infertility cohort, consumption of red meat prior to IVF had a negative influence on embryo development and the likelihood of clinical pregnancy; however, higher fish intake was associated with higher likelihood of blastocyst formation.<sup>96</sup> A case-control study from Hong Kong found that infertile females with unexplained infertility had higher blood mercury concentrations compared to their fertile counterparts.<sup>97</sup> Moreover, higher seafood consumption was associated with elevated blood mercury concentrations in this population. A retrospective time to pregnancy study among pregnant Canadian women found that women with higher mercury concentrations in blood (>1.2 µg/L) or hair (>0.24 ppm) had lower fecundability (FOR=0.22 [95% CI 0.07, 0.72]).<sup>98</sup> A subsequent prospective cohort study of women undergoing IVF in the US, however, did not find any associations between hair mercury levels and any of the intermediate or clinical IVF endpoints.<sup>99</sup> A retrospective cohort study comparing two groups of Swedish women differentially exposed to fatty fish contaminated with persistent organochlorine compounds found no association with time to pregnancy; however, within each group, the consumption of locally caught fatty

fish appeared to have a protective, rather than hazardous, effect on time to pregnancy (success odds ratio=1.27 [95% CI 0.96, 1.69] and 1.36 [95% CI 0.96, 1.94] for women in the east and west coast group, respectively, comparing high to low consumers).<sup>100</sup> In contrast, a retrospective time to pregnancy study among women residing in counties surrounding Lakes Erie and Ontario (two bodies of water with high polychlorinated biphenyls contamination) found that maternal consumption of fish for 3–6 years compared to none was associated with reduced fecundability (conditional FR=0.75 [95% CI 0.51, 0.91]).<sup>101</sup>

Soy, as the main source of phytoestrogens for humans, has received a large degree of attention as a potential reproductive toxicant given well documented and dramatic deleterious reproductive effects due to intake of phytoestrogens initially described in sheep<sup>102</sup> and subsequently identified in other mammals.<sup>103, 104</sup> Evidence from human studies, while limited, has so far not shown little evidence of harm for females. Despite findings from a small study suggesting that soy supplements could improve ovulation,<sup>105</sup> a large cross-sectional analysis with retrospective diet assessment of women participating in the Adventist Health Study found that women with the highest intake of soy isoflavones (~25 times higher than typical intake in Western populations) were 13% (95% CI 2, 26%) more likely to have never been pregnant.<sup>94</sup> However, a prospective cohort study of pregnancy planners in the US found no relation between female urinary isoflavones (a biomarker of soy intake) and fecundity among couples trying to become pregnant (adjusted FORs ranged from 1.02 to 1.05 for a 1 log nmol/L increase in various urinary isoflavones).<sup>106</sup> Furthermore, all published studies evaluating soy intake or phytoestrogen supplements among couples undergoing infertility treatments to date have found them to be beneficial. In a prospective cohort of women undergoing IVF in the US, the odds of achieving a live birth during ART were 77% higher for women with the highest intake of soy isoflavones (mean: 12mg/d; range: 8–28mg/d) than for women who did not consume any soy products.<sup>107</sup> Similarly, isoflavone supplements (120mg/d of isoflavones) increased live birth rates (36.7% versus 13.6%) an RCT among couples undergoing infertility treatment with clomiphene citrate + timed intercourse.<sup>108</sup> Higher doses (1,500mg/d) have also been shown to increase endometrial thickness and ongoing pregnancy rates in women undergoing IUI (20.0% vs. 4.4%)<sup>109</sup> and clinical pregnancy rates in women undergoing IVF (39.3% vs. 20.9%).<sup>110</sup>

At present, there is limited evidence on the association between red or white meat intake and fertility; however the available studies both point to a potential detrimental association between higher red meat intake and higher risk of infertility and adverse embryo development. In regards to fish, the picture is more complicated as the degree of environmental contamination may potentially modify this relationship. Thus, fish from waters with a high degree of environmental pollution as well as those with a high degree of mercury should generally be avoided as the consequences of these environmental toxicants on fertility may outweigh the potential health benefits from the fish alone. Finally, intake of soy supplements and products does not appear to harm fertility as suggested by animal studies, and may in fact confer benefits as suggested by a handful of small studies from infertility cohorts.



## Dietary Patterns

To date, two studies have examined the relation between pre-conception dietary patterns and risk of infertility.<sup>111, 112</sup> In the NHS-II, women in the highest quintile of an investigator-generated “fertility diet” score which prioritizes higher intakes of protein from vegetable sources, full-fat dairy foods, iron, the ratio of MUFAs to trans fats and more frequent use of multivitamins and lower intakes of protein from animal sources, dietary glycemic load, and low fat dairy foods had a 66% (95% CI 52, 77%) lower risk of ovulatory disorder infertility and a 27% (95% CI 5–43%) lower risk of infertility due to other causes compared to women in the lowest quintile.<sup>111</sup> Similarly, a nested case-control study among women in the Seguimiento Universidad de Navarra (SUN) project, found that women with the highest adherence to a Mediterranean-style diet, characterized by higher intakes of vegetables, fruit, fish, poultry, low fat dairy and olive oil, had 0.56 (95% CI 0.35–0.95) times the odds of seeking medical help for difficulty getting pregnant.<sup>112</sup> Two studies from *in vitro* fertilization cohorts further confirm that healthy pre-conception dietary patterns might have a positive impact on fertility.<sup>113, 114</sup> In two separate cohorts, it was shown that higher adherence to the Dutch dietary recommendations (characterized by high intake of whole grains, monounsaturated or polyunsaturated oils, vegetables, fruit, meat or meat replacers, and fish)<sup>114</sup> and higher adherence to a “Mediterranean” diet (characterized by high intake of vegetable oil, fish, legumes, and vegetables and low intake of snacks) prior to treatment was associated with increased probability of pregnancy following IVF (aOR=1.65 [95% CI 1.08, 2.52] for the Dutch dietary pattern and aOR=1.4 [95% CI: 1.0, 1.9] for the Mediterranean dietary pattern).<sup>113</sup> However, despite the increasing evidence suggesting that a healthy pre-conception diet might increase fecundity (or a woman’s chances of becoming pregnant) results from the NHS-II cohort found no relationship between pre-pregnancy adherence to several healthy dietary patterns prior to pregnancy and risk of pregnancy loss.<sup>115</sup>

## Alcohol and caffeine

Intakes of caffeine and alcohol are, unquestionably, the most studied dietary factors as potential disruptors of fertility with more than 30 studies on this topic to date. Results, however, are inconsistent, with multiple studies showing deleterious effects of caffeine<sup>116–125</sup> and alcohol,<sup>126–132</sup> but just as many studies showing no association<sup>125, 126, 133–143</sup><sup>144, 145</sup> or even improved fertility with consumption of certain caffeinated or alcoholic beverages.<sup>134, 139, 146</sup> One potential explanation for these inconsistencies is the fact that most of the studies are retrospective, and thus subject to recall and other types of bias. In fact, systematic reviews on the relation between caffeine and reproductive outcomes have noted that adverse effects of caffeine on reproductive health, including fertility, are more often reported in retrospective studies and studies of low methodological quality;<sup>147, 148</sup> a similar situation may be at play for studies linking alcohol to decreased fertility. While concerns regarding adverse effects of maternal alcohol intake on fetal development are warranted,<sup>149, 150</sup> as are also concerns of increased risk of pregnancy loss with caffeine intake,<sup>39, 151, 152</sup> whether intake of these substances have a deleterious effect on the ability to become pregnant is questionable. In addition, the evidence among couples undergoing ART or other infertility treatments remains relatively slim.<sup>153</sup>

## Paternal Diet

The role of paternal diet on semen quality and couple fertility has been recently examined and summarized in several systematic reviews.<sup>154–157</sup> Some general trends are worth brief mention. First, a Cochrane review of randomized trials of antioxidant supplementation for men in couples undergoing infertility treatment found evidence of benefit for antioxidant supplements in improving semen quality and clinical pregnancy rates.<sup>154</sup> Despite the evidence for benefit, the large heterogeneity of study designs in the trials included in the meta-analysis, along with the expansive definition of “antioxidants” used for the meta-analysis does not make it possible to identify individual agents, combination of agents, or doses responsible for the observed effects. Second, “healthy” dietary patterns (such as the Mediterranean diet pattern and diets characterized by higher intakes of seafood, poultry, whole grains, fruits and vegetables in non-Mediterranean countries) have been consistently associated with better semen parameters, in a wide range of studies in North America, Europe, the Middle East and East Asia.<sup>155, 158</sup> “Unhealthy” diets (rich in red and processed meats, potatoes, sweets and sweetened beverages) have had the opposite relation. Whether these findings can be confirmed in randomized trials remains to be determined. Third, intake of *trans* and saturated fats has consistently been related to poor semen quality;<sup>159–162</sup> *trans* fat intake has also been related to other markers of poor testicular function<sup>163</sup> – including lower testosterone and lower testicular volume – in agreement with animal models.<sup>164–166</sup> Last, moderate intake of alcohol and caffeine do not have a meaningful impact on semen quality.<sup>157, 167</sup> It is important to keep in mind that while much of the research on diet and male fertility has used clinical semen quality parameters as study outcomes, and these remain the cornerstone for the clinical evaluation of the man’s contribution to a couple’s fertility, they are poor predictors of fertility.<sup>168, 169</sup> Hence, associations with semen quality do not imply associations with fertility, and vice versa, as demonstrated by several recent studies among couples undergoing infertility treatment.<sup>170–172</sup>

## Conclusions

The literature on the relation between diet and human fertility has greatly expanded over the last decade and led to the emergence of some clear patterns. Intake of supplemental folic acid has been consistently related to numerous markers of female fertility– from lower frequency of anovulation to higher reproductive success in the setting of ART– suggesting that the reproductive benefits of folate extend beyond the prevention of NTDs. On the other hand, despite promising evidence from animal models, vitamin D does not appear to exert an important role in human fertility in the absence of deficiency. While supplementation with antioxidants does not appear to offer any benefits to women undergoing infertility treatment, it does appear to be beneficial when the male partner is supplemented. However, the available evidence does not allow discerning which specific antioxidants, nor at which doses, are responsible for this benefit. Higher intake of long chain omega 3 fatty acids appears to improve female fertility although it remains unclear whether environmental contamination of fish, their most common food source, can dampen (or even counteract) this benefit. Last, adherence to healthy diets favoring fish, poultry, whole grains, fruits and vegetables, are related to better fertility in women and better semen quality in men. While a complete picture of the role of nutrition on fertility is far from complete, much progress has

been made. Future efforts should concentrate on solidifying emerging evidence and jointly considering female and male diets. Furthermore, to overcome the limitations inherent to observational research based on validated diet assessment tools or nutritional biomarkers, it is essential that the most consistent associations are tested in adequately powered randomized controlled trials.

## Acknowledgments

**Source of Funding:** Supported by NIH grants K99ES026648, R01ES009718 from the NIEHS, L50-HD085359 from the NICHD, and P30DK46200 from the NIDDK

## References

1. Thoma ME, McLain AC, Louis JF, et al. Prevalence of infertility in the United States as estimated by the current duration approach and a traditional constructed approach. *Fertil Steril*. 2013; 99:1324–31. e1. [PubMed: 23290741]
2. Slama R, Hansen OK, Ducot B, et al. Estimation of the frequency of involuntary infertility on a nation-wide basis. *Hum Reprod*. 2012; 27:1489–98. [PubMed: 22416008]
3. Chandra A, Copen CE, Stephen EH. Infertility and impaired fecundity in the United States, 1982–2010: data from the National Survey of Family Growth. *Natl Health Stat Report*. 2013:1–18. 1–19.
4. Assisted reproductive technology in the United States and Canada: 1995 results generated from the American Society for Reproductive Medicine/Society for Assisted Reproductive Technology Registry. *Fertil Steril*. 1998; 69:389–98. [PubMed: 9531864]
5. Preliminary SART Clinic Summary Report: SART (Society for Assisted Reproductive Technologies). 2015; 2017
6. Schieve LA, Devine O, Boyle CA, Petrini JR, Warner L. Estimation of the contribution of non-assisted reproductive technology ovulation stimulation fertility treatments to US singleton and multiple births. *American journal of epidemiology*. 2009; 170:1396–407. [PubMed: 19854803]
7. Chambers GM, Sullivan EA, Ishihara O, Chapman MG, Adamson GD. The economic impact of assisted reproductive technology: a review of selected developed countries. *Fertil Steril*. 2009; 91:2281–94. [PubMed: 19481642]
8. Harris JA, Menke MN, Haefner JK, Moniz MH, Perumalswami CR. Geographic access to assisted reproductive technology health care in the United States: a population-based cross-sectional study. *Fertil Steril*. 2017; 107:1023–27. [PubMed: 28314508]
9. Rossi BV, Bressler LH, Correia KF, Lipskind S, Hornstein MD, Missmer SA. Lifestyle and in vitro fertilization: what do patients believe? *Fertil Res Pract*. 2016; 2:11. [PubMed: 28620538]
10. Showell MG, Brown J, Clarke J, Hart RJ. Antioxidants for female subfertility. *Cochrane Database Syst Rev*. 2013:Cd007807. [PubMed: 23913583]
11. Bibbins-Domingo K, Grossman DC, et al. Force USPST. Folic Acid Supplementation for the Prevention of Neural Tube Defects: US Preventive Services Task Force Recommendation Statement. *JAMA*. 2017; 317:183–89. [PubMed: 28097362]
12. Czeizel AE, Metneki J, Dudas I. The effect of preconceptional multivitamin supplementation on fertility. *Int J Vitam Nutr Res*. 1996; 66:55–8. [PubMed: 8698547]
13. Westphal LM, Polan ML, Trant AS. Double-blind, placebo-controlled study of Fertilityblend: a nutritional supplement for improving fertility in women. *Clin Exp Obstet Gynecol*. 2006; 33:205–8. [PubMed: 17211965]
14. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Use of multivitamins, intake of B vitamins, and risk of ovulatory infertility. *Fertil Steril*. 2008; 89:668–76. [PubMed: 17624345]
15. Gaskins AJ, Mumford SL, Chavarro JE, et al. The impact of dietary folate intake on reproductive function in premenopausal women: a prospective cohort study. *PLoS One*. 2012; 7:e46276. [PubMed: 23050004]
16. Cueto HT, Riis AH, Hatch EE, et al. Folic acid supplementation and fecundability: a Danish prospective cohort study. *Eur J Clin Nutr*. 2016; 70:66–71. [PubMed: 26081493]

17. Thaler CJ, Budiman H, Ruebsamen H, Nagel D, Lohse P. Effects of the common 677C>T mutation of the 5,10-methylenetetrahydrofolate reductase (MTHFR) gene on ovarian responsiveness to recombinant follicle-stimulating hormone. *Am J Reprod Immunol.* 2006; 55:251–8. [PubMed: 1653336]
18. Hecht S, Pavlik R, Lohse P, Noss U, Friese K, Thaler CJ. Common 677C-->T mutation of the 5,10-methylenetetrahydrofolate reductase gene affects follicular estradiol synthesis. *Fertil Steril.* 2009; 91:56–61. [PubMed: 18249399]
19. Szymanski W, Kazdepka-Zieminska A. Effect of homocysteine concentration in follicular fluid on a degree of oocyte maturity. *Ginekol Pol.* 2003; 74:1392–6. [PubMed: 14669450]
20. Gaskins AJ, Afeiche MC, Wright DL, et al. Dietary folate and reproductive success among women undergoing assisted reproduction. *Obstet Gynecol.* 2014; 124:801–9. [PubMed: 25198264]
21. Gaskins AJ, Chiu YH, Williams PL, et al. Association between serum folate and vitamin B-12 and outcomes of assisted reproductive technologies. *Am J Clin Nutr.* 2015; 102:943–50. [PubMed: 26354529]
22. Haggarty P, McCallum H, McBain H, et al. Effect of B vitamins and genetics on success of in-vitro fertilisation: prospective cohort study. *Lancet.* 2006; 367:1513–9. [PubMed: 16679164]
23. Murto T, Kallak TK, Hoas A, et al. Folic acid supplementation and methylenetetrahydrofolate reductase (MTHFR) gene variations in relation to in vitro fertilization pregnancy outcome. *Acta Obstet Gynecol Scand.* 2014
24. Murto T, Skoog Svanberg A, Yngve A, et al. Folic acid supplementation and IVF pregnancy outcome in women with unexplained infertility. *Reprod Biomed Online.* 2014; 28:766–72. [PubMed: 24745837]
25. Ebisch IM, Peters WH, Thomas CM, Wetzels AM, Peer PG, Steegers-Theunissen RP. Homocysteine, glutathione and related thiols affect fertility parameters in the (sub)fertile couple. *Hum Reprod.* 2006; 21:1725–33. [PubMed: 16556671]
26. Boxmeer JC, Macklon NS, Lindemans J, et al. IVF outcomes are associated with biomarkers of the homocysteine pathway in monofollicular fluid. *Hum Reprod.* 2009; 24:1059–66. [PubMed: 19221098]
27. Czeizel AE, Dudas I, Metneki J. Pregnancy outcomes in a randomised controlled trial of periconceptual multivitamin supplementation. Final report *Arch Gynecol Obstet.* 1994; 255:131–9. [PubMed: 7979565]
28. Hook EB, Czeizel AE. Can terathanasia explain the protective effect of folic-acid supplementation on birth defects? *Lancet.* 1997; 350:513–5. [PubMed: 9274597]
29. Windham GC, Shaw GM, Todoroff K, Swan SH. Miscarriage and use of multi-vitamins or folic acid. *Am J Med Genet.* 2000; 90:261–2. [PubMed: 10678670]
30. Wald N, Hackshaw A. Folic acid and prevention of neural-tube defects. *Lancet.* 1997; 350:665.
31. Wald NJ, Hackshaw AK. Folic acid and miscarriage: an unjustified link. *American journal of medical genetics.* 2001; 98:204. [PubMed: 11223859]
32. Central Technical Co-ordinating Unit ITC-oUI. Multicentric study of efficacy of periconceptual folic acid containing vitamin supplementation in prevention of open neural tube defects from India. *Indian J Med Res.* 2000; 112:206–11. [PubMed: 11247198]
33. Balogun OO, da Silva Lopes K, Ota E, et al. Vitamin supplementation for preventing miscarriage. *Cochrane Database Syst Rev.* 2016:CD004073. [PubMed: 27150280]
34. Gindler J, Li Z, Berry RJ, et al. Folic acid supplements during pregnancy and risk of miscarriage. *Lancet.* 2001; 358:796–800. [PubMed: 11564486]
35. Vila-Nova C, Wehby GL, Queiros FC, et al. Periconceptual use of folic acid and risk of miscarriage - findings of the Oral Cleft Prevention Program in Brazil. *Journal of perinatal medicine.* 2013:1–6.
36. Hasan R, Olshan AF, Herring AH, Savitz DA, Siega-Riz AM, Hartmann KE. Self-reported vitamin supplementation in early pregnancy and risk of miscarriage. *Am J Epidemiol.* 2009; 169:1312–8. [PubMed: 19372214]
37. Byrne J. Periconceptual folic acid prevents miscarriage in Irish families with neural tube defects. *Ir J Med Sci.* 2011; 180:59–62. [PubMed: 21052862]

38. Gaskins AJ, Rich-Edwards JW, Hauser R, et al. Maternal prepregnancy folate intake and risk of spontaneous abortion and stillbirth. *Obstet Gynecol.* 2014; 124:23–31. [PubMed: 24901281]
39. Buck Louis GM, Sapra KJ, Schisterman EF, et al. Lifestyle and pregnancy loss in a contemporary cohort of women recruited before conception: The LIFE Study. *Fertil Steril.* 2016; 106:180–8. [PubMed: 27016456]
40. Agic A, Xu H, Altgassen C, et al. Relative expression of 1,25-dihydroxyvitamin D3 receptor, vitamin D 1 alpha-hydroxylase, vitamin D 24-hydroxylase, and vitamin D 25-hydroxylase in endometriosis and gynecologic cancers. *Reproductive sciences (Thousand Oaks, Calif).* 2007; 14:486–97.
41. Parikh G, Varadinova M, Suwandhi P, et al. Vitamin D regulates steroidogenesis and insulin-like growth factor binding protein-1 (IGFBP-1) production in human ovarian cells. *Horm Metab Res.* 2010; 42:754–7. [PubMed: 20711952]
42. Tanamura A, Nomura S, Kurauchi O, Furui T, Mizutani S, Tomoda Y. Purification and characterization of 1,25(OH)2D3 receptor from human placenta. *Journal of obstetrics and gynaecology (Tokyo, Japan).* 1995; 21:631–9.
43. Kwiecinski GG, Petrie GI, DeLuca HF. 1,25-Dihydroxyvitamin D3 restores fertility of vitamin D-deficient female rats. *Am J Physiol.* 1989; 256:E483–7. [PubMed: 2705521]
44. Johnson LE, DeLuca HF. Vitamin D receptor null mutant mice fed high levels of calcium are fertile. *J Nutr.* 2001; 131:1787–91. [PubMed: 11385068]
45. Kovacs CS, Woodland ML, Fudge NJ, Friel JK. The vitamin D receptor is not required for fetal mineral homeostasis or for the regulation of placental calcium transfer in mice. *Am J Physiol Endocrinol Metab.* 2005; 289:E133–44. [PubMed: 15741244]
46. Panda DK, Miao D, Tremblay ML, et al. Targeted ablation of the 25-hydroxyvitamin D 1alpha -hydroxylase enzyme: evidence for skeletal, reproductive, and immune dysfunction. *Proc Natl Acad Sci U S A.* 2001; 98:7498–503. [PubMed: 11416220]
47. Yoshizawa T, Handa Y, Uematsu Y, et al. Mice lacking the vitamin D receptor exhibit impaired bone formation, uterine hypoplasia and growth retardation after weaning. *Nat Genet.* 1997; 16:391–6. [PubMed: 9241280]
48. Mumford SL, Silver R, Sjaarda LA, et al. Vitamin D and Ovarian Reserve and Fecundability among Women with Proven Fecundity. *FASEB J.* 2016; 30(Supplemental):290.6.
49. Moller UK, Streym S, Heickendorff L, Mosekilde L, Rejnmark L. Effects of 25OHD concentrations on chances of pregnancy and pregnancy outcomes: a cohort study in healthy Danish women. *Eur J Clin Nutr.* 2012; 66:862–8. [PubMed: 22378226]
50. Park S, Yoon HK, Ryu HM, et al. Maternal vitamin D deficiency in early pregnancy is not associated with gestational diabetes mellitus development or pregnancy outcomes in Korean pregnant women in a prospective study. *J Nutr Sci Vitaminol (Tokyo).* 2014; 60:269–75. [PubMed: 25297616]
51. Zhou J, Su L, Liu M, et al. Associations between 25-hydroxyvitamin D levels and pregnancy outcomes: a prospective observational study in southern China. *Eur J Clin Nutr.* 2014; 68:925–30. [PubMed: 24865483]
52. Schneuer FJ, Roberts CL, Guilbert C, et al. Effects of maternal serum 25-hydroxyvitamin D concentrations in the first trimester on subsequent pregnancy outcomes in an Australian population. *Am J Clin Nutr.* 2014; 99:287–95. [PubMed: 24257720]
53. Amegah AK, Klever MK, Wagner CL. Maternal vitamin D insufficiency and risk of adverse pregnancy and birth outcomes: A systematic review and meta-analysis of longitudinal studies. *PLoS One.* 2017; 12:e0173605. [PubMed: 28306725]
54. Somigliana E, Paffoni A, Lattuada D, et al. Serum Levels of 25-Hydroxyvitamin D and Time to Natural Pregnancy. *Gynecol Obstet Invest.* 2016; 81:468–71. [PubMed: 26784950]
55. Ozkan S, Jindal S, Greenseid K, et al. Replete vitamin D stores predict reproductive success following in vitro fertilization. *Fertility and sterility.* 2010; 94:1314–9. [PubMed: 19589516]
56. Paffoni A, Ferrari S, Vigano P, et al. Vitamin D deficiency and infertility: insights from in vitro fertilization cycles. *J Clin Endocrinol Metab.* 2014; 99:E2372–6. [PubMed: 25121462]
57. Garbedian K, Boggild M, Moody J, Liu KE. Effect of vitamin D status on clinical pregnancy rates following in vitro fertilization. *CMAJ Open.* 2013; 1:E77–82.



58. Polyzos NP, Anckaert E, Guzman L, et al. Vitamin D deficiency and pregnancy rates in women undergoing single embryo, blastocyst stage, transfer (SET) for IVF/ICSI. *Hum Reprod.* 2014; 29:2032–40. [PubMed: 24951484]
59. Rudick B, Ingles S, Chung K, Stanczyk F, Paulson R, Bendikson K. Characterizing the influence of vitamin D levels on IVF outcomes. *Hum Reprod.* 2012; 27:3321–7. [PubMed: 22914766]
60. Aleyasin A, Hosseini MA, Mahdavi A, et al. Predictive value of the level of vitamin D in follicular fluid on the outcome of assisted reproductive technology. *European journal of obstetrics, gynecology, and reproductive biology.* 2011; 159:132–7.
61. Franasiak JM, Molinaro TA, Dubell EK, et al. Vitamin D levels do not affect IVF outcomes following the transfer of euploid blastocysts. *Am J Obstet Gynecol.* 2015; 212:315e1–6. [PubMed: 25265402]
62. Firouzabadi RD, Rahmani E, Rahsepar M, Firouzabadi MM. Value of follicular fluid vitamin D in predicting the pregnancy rate in an IVF program. *Arch Gynecol Obstet.* 2014; 289:201–6. [PubMed: 23880888]
63. Abadia L, Gaskins AJ, Chiu YH, et al. Serum 25-hydroxyvitamin D concentrations and treatment outcomes of women undergoing assisted reproduction. *Am J Clin Nutr.* 2016; 104:729–35. [PubMed: 27465382]
64. Neville G, Martyn F, Kilbane M, et al. Vitamin D status and fertility outcomes during winter among couples undergoing in vitro fertilization/intracytoplasmic sperm injection. *Int J Gynaecol Obstet.* 2016; 135:172–76. [PubMed: 27530219]
65. Anifandis GM, Dafopoulos K, Messini CI, et al. Prognostic value of follicular fluid 25-OH vitamin D and glucose levels in the IVF outcome. *Reproductive biology and endocrinology: RB&E.* 2010; 8:91. [PubMed: 20667111]
66. Afatoonian A, Arabjahvani F, Eftekhari M, Sayadi M. Effect of vitamin D insufficiency treatment on fertility outcomes in frozen-thawed embryo transfer cycles: A randomized clinical trial. *Iran J Reprod Med.* 2014; 12:595–600. [PubMed: 25469131]
67. Asadi M, Matin N, Frootan M, Mohamadpour J, Qorbani M, Tanha FD. Vitamin D improves endometrial thickness in PCOS women who need intrauterine insemination: a randomized double-blind placebo-controlled trial. *Arch Gynecol Obstet.* 2014; 289:865–70. [PubMed: 24158736]
68. Pal L, Zhang H, Williams J, et al. Vitamin D Status Relates to Reproductive Outcome in Women With Polycystic Ovary Syndrome: Secondary Analysis of a Multicenter Randomized Controlled Trial. *J Clin Endocrinol Metab.* 2016; 101:3027–35. [PubMed: 27186859]
69. Rudick BJ, Ingles SA, Chung K, Stanczyk FZ, Paulson RJ, Bendikson KA. Influence of vitamin D levels on in vitro fertilization outcomes in donor-recipient cycles. *Fertil Steril.* 2014; 101:447–52. [PubMed: 24210230]
70. Fabris A, Pacheco A, Cruz M, Puente JM, Fatemi H, Garcia-Velasco JA. Impact of circulating levels of total and bioavailable serum vitamin D on pregnancy rate in egg donation recipients. *Fertil Steril.* 2014; 102:1608–12. [PubMed: 25256926]
71. Sturmey RG, Reis A, Leese HJ, McEvoy TG. Role of fatty acids in energy provision during oocyte maturation and early embryo development. *Reproduction in domestic animals = Zuchthygiene.* 2009; 44(Suppl 3):50–8. [PubMed: 19660080]
72. Norwitz ER, Schust DJ, Fisher SJ. Implantation and the survival of early pregnancy. *N Engl J Med.* 2001; 345:1400–8. [PubMed: 11794174]
73. Nehra D, Le HD, Fallon EM, et al. Prolonging the female reproductive lifespan and improving egg quality with dietary omega-3 fatty acids. *Aging cell.* 2012; 11:1046–54. [PubMed: 22978268]
74. Hammiche F, Vujkovic M, Wijburg W, et al. Increased preconception omega-3 polyunsaturated fatty acid intake improves embryo morphology. *Fertil Steril.* 2011; 95:1820–3. [PubMed: 21130435]
75. Lefevre M, Lovejoy JC, Smith SR, et al. Comparison of the acute response to meals enriched with cis- or trans-fatty acids on glucose and lipids in overweight individuals with differing FABP2 genotypes. *Metabolism: clinical and experimental.* 2005; 54:1652–8. [PubMed: 16311100]
76. Kaipia A, Chun SY, Eisenhauer K, Hsueh AJ. Tumor necrosis factor-alpha and its second messenger, ceramide, stimulate apoptosis in cultured ovarian follicles. *Endocrinology.* 1996; 137:4864–70. [PubMed: 8895358]



77. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Dietary fatty acid intakes and the risk of ovulatory infertility. *The American journal of clinical nutrition*. 2007; 85:231–7. [PubMed: 17209201]
78. Wise LA, Wesselink AK, Tucker KL, et al. Dietary fat intake and fecundability in two preconception cohort studies. *Am J Epidemiol*. 2017
79. Mumford SL, Chavarro JE, Zhang C, et al. Dietary fat intake and reproductive hormone concentrations and ovulation in regularly menstruating women. *Am J Clin Nutr*. 2016; 103:868–77. [PubMed: 26843151]
80. Moran LJ, Tzagareli V, Noakes M, Norman R. Altered Preconception Fatty Acid Intake Is Associated with Improved Pregnancy Rates in Overweight and Obese Women Undertaking In Vitro Fertilisation. *Nutrients*. 2016:8.
81. Jungheim ES, Frolova AI, Jiang H, Riley JK. Relationship between serum polyunsaturated fatty acids and pregnancy in women undergoing in vitro fertilization. *J Clin Endocrinol Metab*. 2013; 98:E1364–8. [PubMed: 23780371]
82. Mirabi P, Chaichi MJ, Esmaeilzadeh S, et al. The role of fatty acids on ICSI outcomes: a prospective cohort study. *Lipids Health Dis*. 2017; 16:18. [PubMed: 28109274]
83. Swarts WJ, Mattison DR. Galactose inhibition of ovulation in mice. *Fertil Steril*. 1988; 49:522–26. [PubMed: 3342905]
84. Bandyopadhyay S, Chakrabarti J, Banerjee S, et al. Galactose toxicity in the rat as a model for premature ovarian failure: an experimental approach readdressed. *Hum Reprod*. 2003; 18:2031–8. [PubMed: 14507817]
85. Garcia-Pelaez B, Ferrer-Lorente R, Gomez-Olles S, Fernandez-Lopez JA, Remesar X, Alemany M. Technical note: Measurement of total estrone content in foods. Application to dairy products. *J Dairy Sci*. 2004; 87:2331–6. [PubMed: 15328253]
86. Cramer DW, Xu H, Sahi T. Adult hypolactasia, milk consumption, and age-specific fertility. *Am J Epidemiol*. 1994; 139:282–9. [PubMed: 8116603]
87. Greenlee AR, Arbuckle TE, Chyou PH. Risk factors for female infertility in an agricultural region. *Epidemiology*. 2003; 14:429–36. [PubMed: 12843768]
88. Chavarro JE, Rich-Edwards JW, Rosner B, Willett WC. A prospective study of dairy foods intake and anovulatory infertility. *Human reproduction*. 2007; 22:1340–7. [PubMed: 17329264]
89. Afeiche MC, Chiu YH, Gaskins AJ, et al. Dairy intake in relation to in vitro fertilization outcomes among women from a fertility clinic. *Hum Reprod*. 2016
90. Wise LA, Wesselink AK, Mikkelsen EM, et al. Dairy intake and fecundability in 2 preconception cohort studies. *Am J Clin Nutr*. 2017; 105:100–10. [PubMed: 27903519]
91. Jeong SH, Kang D, Lim MW, Kang CS, Sung HJ. Risk assessment of growth hormones and antimicrobial residues in meat. *Toxicological research*. 2010; 26:301–13. [PubMed: 24278538]
92. Fraser AJ, Webster TF, McClean MD. Diet contributes significantly to the body burden of PBDEs in the general U.S. population. *Environ Health Perspect*. 2009; 117:1520–5. [PubMed: 20019900]
93. Vandermeersch G, Lourenco HM, Alvarez-Munoz D, et al. Environmental contaminants of emerging concern in seafood—European database on contaminant levels. *Environ Res*. 2015; 143:29–45. [PubMed: 26123540]
94. Jacobsen BK, Jaceldo-Siegl K, Knutsen SF, Fan J, Oda K, Fraser GE. Soy isoflavone intake and the likelihood of ever becoming a mother: the Adventist Health Study-2. *International journal of women's health*. 2014; 6:377–84.
95. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Protein intake and ovulatory infertility. *Am J Obstet Gynecol*. 2008; 198:210e1–7. [PubMed: 18226626]
96. Braga DP, Halpern G, Setti AS, Figueira RC, Iaconelli A Jr, Borges E Jr. The impact of food intake and social habits on embryo quality and the likelihood of blastocyst formation. *Reprod Biomed Online*. 2015; 31:30–8. [PubMed: 25982093]
97. Choy CM, Lam CW, Cheung LT, Briton-Jones CM, Cheung LP, Haines CJ. Infertility, blood mercury concentrations and dietary seafood consumption: a case-control study. *BJOG*. 2002; 109:1121–5. [PubMed: 12387464]

98. Cole DC, Wainman B, Sanin LH, Weber JP, Muggah H, Ibrahim S. Environmental contaminant levels and fecundability among non-smoking couples. *Reprod Toxicol.* 2006; 22:13–9. [PubMed: 16439098]
99. Wright DL, Afeiche MC, Ehrlich S, et al. Hair mercury concentrations and in vitro fertilization (IVF) outcomes among women from a fertility clinic. *Reprod Toxicol.* 2015; 51:125–32. [PubMed: 25601638]
100. Axmon A, Rylander L, Stromberg U, Hagmar L. Female fertility in relation to the consumption of fish contaminated with persistent organochlorine compounds. *Scand J Work Environ Health.* 2002; 28:124–32. [PubMed: 12019589]
101. Buck GM, Vena JE, Schisterman EF, et al. Parental consumption of contaminated sport fish from Lake Ontario and predicted fecundability. *Epidemiology.* 2000; 11:388–93. [PubMed: 10874544]
102. Bennetts HW, Underwood EJ, Shier FL. A specific breeding problem of sheep on subterranean clover pastures in Western Australia. *Austral Vet J.* 1946; 22:2–12. [PubMed: 21028682]
103. Seppen J. A diet containing the soy phytoestrogen genistein causes infertility in female rats partially deficient in UDP glucuronyltransferase. *Toxicol Appl Pharmacol.* 2012; 264:335–42. [PubMed: 23000043]
104. Setchell KD, Gosselin SJ, Welsh MB, et al. Dietary estrogens--a probable cause of infertility and liver disease in captive cheetahs. *Gastroenterology.* 1987; 93:225–33. [PubMed: 3297906]
105. Kohama T, Kobayashi H, Inoue M. The effect of soybeans on the anovulatory cycle. *J Med Food.* 2005; 8:550–1. [PubMed: 16379571]
106. Mumford SL, Sundaram R, Schisterman EF, et al. Higher urinary lignan concentrations in women but not men are positively associated with shorter time to pregnancy. *J Nutr.* 2014; 144:352–8. [PubMed: 24401816]
107. Vanegas JC, Afeiche MC, Gaskins AJ, et al. Soy food intake and treatment outcomes of women undergoing assisted reproductive technology. *Fertil Steril.* 2015; 103:749–55. e2. [PubMed: 25577465]
108. Shahin AY, Ismail AM, Zahran KM, Makhlof AM. Adding phytoestrogens to clomiphene induction in unexplained infertility patients--a randomized trial. *Reprod Biomed Online.* 2008; 16:580–88. [PubMed: 18413068]
109. Unfer V, Casini ML, Costabile L, Mignosa M, Gerli S, Di Renzo GC. High dose of phytoestrogens can reverse the antiestrogenic effects of clomiphene citrate on the endometrium in patients undergoing intrauterine insemination: a randomized trial. *J Soc Gynecol Investig.* 2004; 11:323–8.
110. Unfer V, Casini ML, Gerli S, Costabile L, Mignosa M, Di Renzo GC. Phytoestrogens may improve the pregnancy rate in in vitro fertilization-embryo transfer cycles: a prospective, controlled, randomized trial. *Fertil Steril.* 2004; 82:1509–13. [PubMed: 15589851]
111. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Diet and lifestyle in the prevention of ovulatory disorder infertility. *Obstetrics and gynecology.* 2007; 110:1050–8. [PubMed: 17978119]
112. Toledo E, Lopez-del Burgo C, Ruiz-Zambrana A, et al. Dietary patterns and difficulty conceiving: a nested case-control study. *Fertility and sterility.* 2011; 96:1149–53. [PubMed: 21943725]
113. Vujkovic M, de Vries JH, Lindemans J, et al. The preconception Mediterranean dietary pattern in couples undergoing in vitro fertilization/intracytoplasmic sperm injection treatment increases the chance of pregnancy. *Fertil Steril.* 2010; 94:2096–101. [PubMed: 20189169]
114. Twigt JM, Bolhuis ME, Steegers EA, et al. The preconception diet is associated with the chance of ongoing pregnancy in women undergoing IVF/ICSI treatment. *Hum Reprod.* 2012; 27:2526–31. [PubMed: 22593431]
115. Gaskins AJ, Rich-Edwards JW, Hauser R, et al. Prepregnancy dietary patterns and risk of pregnancy loss. *Am J Clin Nutr.* 2014; 100:1166–72. [PubMed: 25240079]
116. Wilcox A, Weinberg C, Baird D. Caffeinated beverages and decreased fertility. *Lancet.* 1988; 332:1453–56.
117. Christianson R, Oechsli F, Van Den Berg B. Caffeinated beverages and decreased fertility. *Lancet.* 1989; 333:378.

118. Williams M, Monson R, Goldman M, Mittendorf R, Ryan K. Coffee and delayed conception. *Lancet*. 1990; 335:1603.
119. Hatch EE, Bracken MB. Association of Delayed Conception with Caffeine Consumption. *Am J Epidemiol*. 1993; 138:1082–92. [PubMed: 8266910]
120. Bolumar F, Olsen J, Rebagliato M, Bisanti L. Subfecundity ESGoI. Caffeine intake and delayed conception: a European multicenter study on infertility and subfecundity. *Am J Epidemiol*. 1997; 145:324–34. [PubMed: 9054236]
121. Stanton CK, Gray RH. Effects of Caffeine Consumption on Delayed Conception. *Am J Epidemiol*. 1995; 142:1322–29. [PubMed: 7503053]
122. Jensen TK, Henriksen TB, Hjollund NHI, et al. Caffeine Intake and Fecundability: A Follow-up Study among 430 Danish Couples Planning Their First Pregnancy. *Reprod Toxicol*. 1998; 12:289–95. [PubMed: 9628552]
123. Grodstein F, Goldman MB, Ryan L, Cramer DW. Relation of Female Infertility to Consumption of Caffeinated Beverages. *Am J Epidemiol*. 1993; 137:1353–60. [PubMed: 8333416]
124. Olsen J. Cigarette smoking, tea and coffee drinking, and subfecundity. *Am J Epidemiol*. 1991; 133:734–39. [PubMed: 2018028]
125. Hassan MAM, Killick SR. Negative lifestyle is associated with a significant reduction in fecundity. *Fertil Steril*. 2004; 81:384–92. [PubMed: 14967378]
126. Hakim RB, Gray RH, Zacur H. Alcohol and caffeine consumption and decreased fertility. *Fertil Steril*. 1998; 70:632–37. [PubMed: 9797089]
127. Jensen TK, Hjollund NHI, Henriksen TB, et al. Does moderate alcohol consumption affect fertility? Follow up study among couples planning first pregnancy. *Br Med J*. 1998; 317:505–10. [PubMed: 9712595]
128. Eggert J, Theobald H, Engfeldt P. Effects of alcohol consumption on female fertility during and 18-year period. *Fertil Steril*. 2004; 81:379–83. [PubMed: 14967377]
129. Greenlee AR, Arbuckle TE, Chyou PH. Risk factors for female infertility in an agricultural region. *Epidemiology*. 2003; 14:429–36. [PubMed: 12843768]
130. Idrovo AJ, Sanin LH, Cole D, et al. Time to first pregnancy among women working in agricultural production. *Int Arch Occup Environ Health*. 2005; 78:493–500. [PubMed: 15918035]
131. Tolstrup JS, Kjaeger SK, Holst C, et al. Alcohol use as a predictor for infertility in a representative population of Danish women. *Acta Obstet Gynecol Scand*. 2003; 82:744–49. [PubMed: 12848646]
132. Grodstein F, Goldman MB, Cramer DW. Infertility in women and moderate alcohol consumption. *Am J Public Health*. 1994; 85:1021–22.
133. Olsen J, Rachootin P, Schiodt AV, Damsbo N. Tobacco Use, Alcohol Consumption and Infertility. *Int J Epidemiol*. 1983; 12:179–84. [PubMed: 6874213]
134. Florack EIM, Zielhuis GA, Rolland R. Cigarette Smoking, Alcohol Consumption, and Caffeine Intake and Fecundability. *Prev Med*. 1994; 23:175–80. [PubMed: 8047523]
135. Curtis KM, Savitz DA, Arbuckle TE. Effects of Cigarette Smoking, Caffeine Consumption, and Alcohol Intake on Fecundability. *Am J Epidemiol*. 1997; 146:32–41. [PubMed: 9215221]
136. Olsen J, Bolumar F, Boldsen J, Bisanti L. Does Moderate Alcohol Intake Reduce Fecundability? A European Multicenter Study on Infertility and Subfecundity. *Alcohol Clin Exp Res*. 1997; 21:206–12. [PubMed: 9113254]
137. Parazzini F, Chatenoud L, Di Cintio E, La Vecchia C, Benzi G, Fedele L. Alcohol consumption is not related to fertility in Italian women. *Br Med J*. 1999; 318:397.
138. Juhl M, Andersen AMN, Gronbaek M, Olsen J. Moderate alcohol consumption and waiting time to pregnancy. *Hum Reprod*. 2002; 16:2705–09.
139. Spinelli A, Figa-Talamanca I, Osborn J. Time to pregnancy and occupation in a group of Italian women. *Int J Epidemiol*. 1997; 26:601–07. [PubMed: 9222786]
140. Joesoef MR, Beral V, Rolfs RT, Aral SO, Cramer DW. Are caffeinated beverages risk factors for delayed conception? *Lancet*. 1990; 335:136–37. [PubMed: 1967434]

141. Alderete E, Eskenazi B, Sholtz R. Effect of Cigarette Smoking and Coffee Drinking on Time to Conception. *Epidemiology*. 1995; 6:403–08. [PubMed: 7548349]
142. Caan B, Quesenberry CP Jr, Coates AO. Differences in fertility associated with caffeinated beverage consumption. *Am J Public Health*. 1998; 88:270–74. [PubMed: 9491020]
143. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Caffeinated and alcoholic beverage intake in relation to ovulatory disorder infertility. *Epidemiology*. 2009; 20:374–81. [PubMed: 19279491]
144. Mikkelsen EM, Riis AH, Wise LA, et al. Alcohol consumption and fecundability: prospective Danish cohort study. *Bmj*. 2016; 354:i4262. [PubMed: 27581754]
145. Hatch EE, Wise LA, Mikkelsen EM, et al. Caffeinated beverage and soda consumption and time to pregnancy. *Epidemiology*. 2012; 23:393–401. [PubMed: 22407137]
146. Juhl M, Olsen J, Andersen AMN, Gronbaek M. Intake of wine, beer and spirits and waiting time to pregnancy. *Hum Reprod*. 2003; 18:1967–71. [PubMed: 12923158]
147. Peck JD, Leviton A, Cowan LD. A review of the epidemiologic evidence concerning the reproductive health effects of caffeine consumption: a 2000–2009 update. *Food Chem Toxicol*. 2010; 48:2549–76. [PubMed: 20558227]
148. Leviton A, Cowan L. A review of the literature relating caffeine consumption by women to their risk of reproductive hazards. *Food Chem Toxicol*. 2002; 40:1271–310. [PubMed: 12204391]
149. Williams JF, Smith VC. Fetal Alcohol Spectrum Disorders. *Pediatrics*. 2015; 136:e1395–406. [PubMed: 26482673]
150. Dorrie N, Focker M, Freunschit I, Hebebrand J. Fetal alcohol spectrum disorders. *European child & adolescent psychiatry*. 2014; 23:863–75. [PubMed: 24965796]
151. Greenwood DC, Thatcher NJ, Ye J, et al. Caffeine intake during pregnancy and adverse birth outcomes: a systematic review and dose-response meta-analysis. *Eur J Epidemiol*. 2014; 29:725–34. [PubMed: 25179792]
152. Gaskins AJ, Rich-Edwards JW, Williams PL, Toth TL, Missmer SA, Chavarro JE. Pre-pregnancy caffeine and caffeinated beverage intake and risk of spontaneous abortion. *Eur J Nutr*. 2016
153. Abadia L, Chiu YH, Williams PL, et al. The association between pre-treatment maternal alcohol and caffeine intake and outcomes of assisted reproduction in a prospectively followed cohort. *Hum Reprod*. 2017 In Press.
154. Showell MG, Mackenzie-Proctor R, Brown J, Yazdani A, Stankiewicz MT, Hart RJ. Antioxidants for male subfertility. *Cochrane Database Syst Rev*. 2014:Cd007411. [PubMed: 25504418]
155. Salas-Huetos A, Bullo M, Salas-Salvado J. Dietary patterns, foods and nutrients in male fertility parameters and fecundability: a systematic review of observational studies. *Hum Reprod Update*. 2017:1–19. [PubMed: 29190351]
156. Giahi L, Mohammadmoradi S, Javidan A, Sadeghi MR. Nutritional modifications in male infertility: a systematic review covering 2 decades. *Nutr Rev*. 2016; 74:118–30. [PubMed: 26705308]
157. Li Y, Lin H, Li Y, Cao J. Association between socio-psycho-behavioral factors and male semen quality: systematic review and meta-analyses. *Fertil Steril*. 2011; 95:116–23. [PubMed: 20674912]
158. Liu CY, Chou YC, Chao JC, Hsu CY, Cha TL, Tsao CW. The Association between Dietary Patterns and Semen Quality in a General Asian Population of 7282 Males. *PLoS One*. 2015; 10:e0134224. [PubMed: 26218796]
159. Jensen TK, Heitmann BL, Jensen MB, et al. High dietary intake of saturated fat is associated with reduced semen quality among 701 young Danish men from the general population. *Am J Clin Nutr*. 2013; 97:411–8. [PubMed: 23269819]
160. Attaman JA, Toth TL, Furtado J, Campos H, Hauser R, Chavarro JE. Dietary fat and semen quality among men attending a fertility clinic. *Hum Reprod*. 2012; 27:1466–74. [PubMed: 22416013]
161. Chavarro JE, Minguéz-Alarcon L, Mendiola J, Cutillas-Tolin A, Lopez-Espin JJ, Torres-Cantero AM. Trans fatty acid intake is inversely related to total sperm count in young healthy men. *Hum Reprod*. 2014; 29:429–40. [PubMed: 24419496]

162. Chavarro JE, Furtado J, Toth TL, et al. Trans-fatty acid levels in sperm are associated with sperm concentration among men from an infertility clinic. *Fertil Steril*. 2011; 95:1794–7. [PubMed: 21071027]
163. Minguez-Alarcon L, Chavarro JE, Mendiola J, et al. Fatty acid intake in relation to reproductive hormones and testicular volume among young healthy men. *Asian J Androl*. 2017; 19:184–90. [PubMed: 27834316]
164. Jensen B. Rat testicular lipids and dietary isomeric fatty acids in essential fatty acid deficiency. *Lipids*. 1976; 11:179–88. [PubMed: 1263760]
165. Hanis T, Zidek V, Sachova J, Klir P, Deyl Z. Effects of dietary trans-fatty acids on reproductive performance of Wistar rats. *British Journal of Nutrition*. 1989; 61:519–29. [PubMed: 2758008]
166. Veaute C, Andreoli MF, Racca A, et al. Effects of Isomeric Fatty Acids on Reproductive Parameters in Mice. *Am J Reprod Immunol*. 2007; 58:487–96. [PubMed: 17997747]
167. Jensen TK, Swan S, Jorgensen N, et al. Alcohol and male reproductive health: a cross-sectional study of 8344 healthy men from Europe and the USA. *Hum Reprod*. 2014; 29:1801–9. [PubMed: 24893607]
168. Buck Louis GM, Sundaram R, Schisterman EF, et al. Semen quality and time to pregnancy: the Longitudinal Investigation of Fertility and the Environment Study. *Fertil Steril*. 2014; 101:453–62. [PubMed: 24239161]
169. Patel CJ, Sundaram R, Buck Louis GM. A data-driven search for semen-related phenotypes in conception delay. *Andrology*. 2017; 5:95–102. [PubMed: 27792860]
170. Xia W, Chiu YH, Williams PL, et al. Men’s meat intake and treatment outcomes among couples undergoing assisted reproduction. *Fertil Steril*. 2015; 104:972–9. [PubMed: 26206344]
171. Karmon AE, Toth TL, Chiu YH, et al. Male caffeine and alcohol intake in relation to semen parameters and in vitro fertilization outcomes among fertility patients. *Andrology*. 2017
172. Minguez-Alarcon L, Afeiche MC, Chiu YH, et al. Male soy food intake was not associated with in vitro fertilization outcomes among couples attending a fertility center. *Andrology*. 2015; 3:702–8. [PubMed: 26097060]

**Table 1**

Overview of the literature on the relation between diet and female fertility

	What is the bottom line?	What are the gaps in the evidence?	I want to read more but do not have much time.
Antioxidant supplements	Antioxidant supplements most likely do not make a difference	Too few studies have tested the exact same intervention so it is difficult to draw strong conclusions	Ref. 10
Folic acid, vitamin B12	Folic acid may increase fertility and live birth rates in ART. Doses higher than recommended for NTD prevention may offer the greatest benefit as might the additional intake of vitamin B12	No randomized trials have tested the doses related with greatest benefit in observational studies	Ref. 16 and 20 or 21
Vitamin D	Vitamin D does not have a major impact on fertility within the observed range of supplementation/adequate serum levels	Most published work has focused on women with vitamin D intakes or serum concentrations within or very close to normal range; an adverse effect of severe deficiency on fertility cannot be ruled out	Ref. 53 and 61 or 63
Dietary fats	<i>Trans</i> fatty acids (even at current intake levels in the US) are related to lower fertility while long chain omega-3 fatty acids have the opposite relation	The role of <i>trans</i> fatty acids will become an untestable hypothesis as they are phased out of the US food supply, but they may still be relevant elsewhere Trials of omega-3 fatty acid supplementation are needed	Ref 77, 78, 81, 82
Dairy	Dairy foods probably do not have an important influence on fertility	Very few studies have addressed this question	Ref 89, 90
Meats	Intake of red meats and fish with high levels of environmental contamination may be of concern	Very few studies have addressed this question	Ref 95, 96, 101
Soy, isoflavones	Soy intake does not help or hurt couples trying to conceive on their own; however, isoflavone intake may increase live birth rates in ART	Only one study to date among pregnancy planners Vast range of doses in ART studies yet all show similar effects	Ref. 106, 110
Diet patterns	“Healthy” diets have been consistently related to better fertility and higher live birth rates in ART across multiple studies. “Unhealthy” diets have consistently had the opposite relation.	Definition of healthy and unhealthy diets changes slightly from study to study. No randomized trials to date.	Ref. 111, 113
Alcohol, caffeine	Most large, well designed studies have not detected associations between higher alcohol or caffeine intake and lower fertility	Since randomized trials of alcohol/caffeine will likely be judged as unethical, there is a need for more large, high quality prospective cohort studies to clarify this issue.	Ref 144, 145, 153



Overview of the literature on the relation between diet and male fertility

Table 2

	What is the bottom line?	What are the gaps in the evidence?	I want to read more but do not have much time.
Antioxidant supplements	Supplementing men in couples undergoing ART with antioxidants may increase live birth rates	Based on the current literature, it is not possible to say, what antioxidants (or combinations) or at what doses are responsible for this benefit.	Ref 154
Diet patterns	“Healthy” diets have been consistently related to better semen quality across a wide range of populations. “Unhealthy” diets have consistently had the opposite relation	Definitions of healthy and unhealthy diets change slightly from study to study. No randomized trials to date. Effect on semen quality does not imply effect on couple fertility.	Ref 155
Dietary fats	Intake of saturated and <i>trans</i> fats has consistently been related to lower semen quality and other markers of poor testicular function.	No randomized trials to date. Effect on semen quality does not imply effect on couple fertility.	Ref 159, 161, 163
Alcohol, caffeine	Alcohol and caffeine do not have an important impact on semen quality within usual ranges of intake. The exception is alcohol intake at levels associated with liver disease.	Effect on semen quality does not imply effect on couple fertility.	Ref 157