

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/6510449>

Efficacy of α -Lipoic Acid Against Diabetic Cataract in Rat

Article in *Japanese Journal of Ophthalmology* · February 2007

DOI: 10.1007/s10384-006-0384-3 · Source: PubMed

CITATIONS

29

READS

223

6 authors, including:



Hiroshi Sasaki

Kanazawa Medical University

172 PUBLICATIONS 2,606 CITATIONS

SEE PROFILE

Some of the authors of this publication are also working on these related projects:



5-S-GAD [View project](#)

LABORATORY INVESTIGATION

Efficacy of α -Lipoic Acid Against Diabetic Cataract in Rat

Masami Kojima^{1,2}, Li Sun², Ikuho Hata², Yasuo Sakamoto^{1,2}, Hiroshi Sasaki^{1,2},
and Kazuyuki Sasaki²

¹Department of Ophthalmology, Kanazawa Medical University, Ishikawa, Japan;

²Division of Vision Research for Environmental Health, Medical Research Institute,
Kanazawa Medical University, Ishikawa, Japan

Abstract

Purpose: α -Lipoic acid (LA) is well known as a powerful antioxidant. The efficacy of dihydrolipoate-LA for oral administration against streptozotocin (STZ)-induced diabetic cataract in rat was investigated.

Methods: Rats were divided into three groups, control, diabetes mellitus (DM), and DM treated with LA (DM+LA). Diabetes was induced by intravenous injection of 50 mg/kg STZ. DM+LA rats were fed 30 mg/rat per day LA in their diet. Lens changes were assessed using Scheimpflug images (EAS-1000) and by measuring light-scattering intensity.

Results: Increase in lens light scattering was less ($P < 0.05$) in DM+LA rats than in DM rats 5 weeks after induction of diabetes. DM rats had the highest and control rats the lowest blood glucose levels at every measurement point up to 111 days ($P < 0.05$).

Conclusion: LA treatment delayed development and progression of cataract in rats with streptozotocin-induced diabetes. **Jpn J Ophthalmol** 2007;51:10-13 © Japanese Ophthalmological Society 2007

Key Words: antioxidant, diabetic cataract, α -lipoic acid, lipoic acid, rat

Introduction

α -Lipoic acid (LA) and its reduced form, dihydrolipoate (DHLA), are both powerful antioxidants. Both act not only directly, by radical quenching and metal chelation, but also indirectly, through the recycling of other antioxidants such as ascorbic acid, vitamin E, and glutathione.^{1,2} LA and DHLA have received much attention because many diabetic complications are believed to be mediated by oxygen free-radical generation. LA has been shown to protect against cataractogenesis, a diabetic complication resulting from polyol accumulation, crystallin glycation, and oxidative insults,³ in both in vivo and in vitro models.⁴⁻¹⁰

LA prevented cataract formation by increasing glutathione, ascorbate, and vitamin E levels, and restoring the

activities of glutathione, peroxidase, catalase, and ascorbate free-radical reductase in the lenses of 60% of rats treated with L-buthionine sulfoximine, an inhibitor of glutathione synthesis.⁴ Intraperitoneal injection of LA significantly reduced blood glucose levels, increased lenticular glutathione content, and inhibited cataractogenesis in a sand rat model of acute type 2 diabetes.⁹ However, LA delivered by intubation had no effect on blood glucose levels and cataract development in sand rats fed a "medium-energy" diet (a chronic diabetic model), although glutathione levels increased.⁹ In addition, it has been reported that LA might prevent protein glycation¹¹ and inhibit aldose reductase activity in lenses cultured under hyperglycemic conditions.¹²

Although several reports have shown that LA may prevent formation of experimentally induced cataract, it remains unclear whether LA can also prevent or delay cataract progression. Therefore, to investigate the effects of LA on cataract progression in streptozotocin-induced diabetic rats over a 10-month period, we periodically analyzed photographs of the lenses.

Received: May 22, 2006 / Accepted: September 4, 2006

Correspondence and reprint requests to: Masami Kojima, Department of Ophthalmology, Kanazawa Medical University, 1-1 Daigaku, Uchinada-machi, Kahoku-gun, Ishikawa 920-0293, Japan
e-mail: m-kojima@kanazawa-med.ac.jp

Table 1. Blood glucose levels

Blood glucose (mg/dl)	Days after DM induction			
	3 days	41 days	80 days	111 days
CTL	80.7 ± 3.9*	82.6 ± 8.1	72.5 ± 2.1*	—
DM	381.1 ± 28.8	>500	686.2 ± 81.2	620.9 ± 111.2
DM+LA	347.8 ± 70.5	369.1 ± 72.4	531.4 ± 168.1*	514.8 ± 99.7*

CTL, control; DM, diabetes mellitus; LA, α -lipoic acid.* $P < 0.05$ vs. DM.

Materials and Methods

Thirty-six 7-week-old female Brown Norway rats were used in this study. All animals were cared for and handled in accordance with Guidelines for Animal Experiments at Kanazawa Medical University and the ARVO Resolution on the Use of Animals in Research. Food and water were provided ad libitum.

Rats were divided into control (CTL, 8 rats), diabetes mellitus (DM, 14 rats), and DM with LA treatment groups (DM+LA, 14 rats). Diabetes was induced by intravenous injection of 50 mg/kg streptozotocin (STZ) (2% solution in 0.1 M citrate buffer, pH 4.6), and onset was confirmed in all rats 3 days later by blood glucose levels exceeding 200 mg/dl (11 mmol/l), as measured with a Glutest GT-1610 analyzer (Sanwa Kagaku Kenkyusho, Nagoya, Japan).

The same volume of citrate buffer without STZ was injected into the CTL rats. LA (Wako Pure Chemical, Osaka, Japan, 30 mg/rat per day) was mixed with powdered food and given as a daily diet supplement to the DM+LA group starting 3 days after diabetes induction. The mixture rate of the LA in the food had been properly changed from the food intake the day before. Lenses were photographed weekly using an anterior eye segment system (EAS-1000, NIDEK, Gamagori, Japan). Light-scattering intensities at 0.08, 0.13, 0.18, and 0.23 mm from the center of the anterior capsule were measured to evaluate the extent of lens opacification as described by Kojima and Sasaki.¹³ Blood glucose levels were measured at 3 and 41 days after diabetes induction with a glucose measurement kit (Glutest Sensor, Sanwa), 80 and 111 days after diabetes induction (Glucose Test, Wako). Food consumption and body weight were measured daily. The data are expressed as means \pm SD. Differences between groups were assessed using the Student *t* test, and *P* values less than 0.05 were considered to be significant.

Results

In the CTL group, mean body weight increased from 100 to 180 g during the first 3 months, and then gradually leveled off at 200 g at 5 months. In contrast, mean body weight did not change in the DM or DM+LA groups, remaining at around 100 g. There were differences ($P < 0.01$) in body weight between the CTL and DM groups, and also between

the CTL and DM+LA groups from the fourth day onward. Treating DM rats with LA had no significant effect (DM vs. DM+LA) on weight gain. The average food consumption was highest in the DM group (13.4 g/rat per day) followed by the CTL (11.4 g/rat per day) and DM+LA groups (9.8 g/rat per day). Significant differences were found among the groups ($P < 0.05$). Blood glucose levels were highest in the DM group at every measurement point, and an inhibitory effect on elevation of blood glucose was observed in the DM+LA group (Table 1). Significant differences in blood glucose levels were observed between CTL and DM groups, and between CTL and DM+LA groups at some measurement points ($P < 0.05$ in both cases). Blood glucose levels on day 80 and 111 were significantly lower ($P < 0.05$, $P < 0.01$ respectively) in the DM+LA groups than in the DM group.

Figure 1 shows representative Scheimpflug slit images for each group. A slight increase in opacity in the anterior subcapsular region was first observed 3 weeks after diabetes induction in the DM group, and the opacity subsequently became denser and spread over a larger area. In the DM+LA group, the development and progression of the opacity was much slower. Figure 2 shows changes in light-scattering intensity with time at a depth of 0.08 mm from the center of the anterior capsule. A rapid increase in light-scattering intensity was observed in the DM group starting from 3 weeks after the experimental induction of diabetes. Although the DM+LA group showed the same pattern of change as the DM group, the pace of the increase in the DM+LA group was much more gradual. Significant differences were found between the DM and CTL groups and also between the DM and DM+LA groups from 3 weeks. A significant difference between the CTL and DM+LA groups was found from 5 weeks. Increases in light-scattering intensity measured at deeper points (0.13, 0.18, and 0.23 mm depth from the anterior capsule) were detected later than increases at shallower points (data not shown).

During the process of cataract development in STZ-induced diabetic rats, initial signs of lens opacification have been reported to appear in the anterior subcapsular region, followed by the anterior deep cortex, exhibiting a diffused cloudiness that can be distinguished from the relatively transparent superficial cortex. This can explain the decrease of lens light-scattering intensity in both the DM and DM+LA groups at a depth of 0.08 mm at 9 weeks and thereafter.

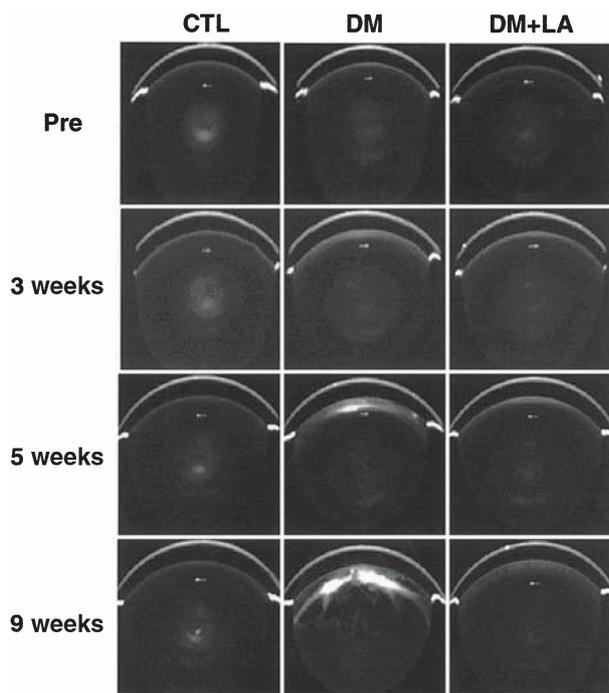


Figure 1. Scheimpflug slit images for each group. Slight changes around the anterior subcapsular region were observed 3 weeks after diabetes induction in the diabetes mellitus (DM) group. The opacities in the DM group rapidly enlarged and increased in density, while those in the DM with α -lipoic acid treatment (DM+LA) group progressed more slowly.

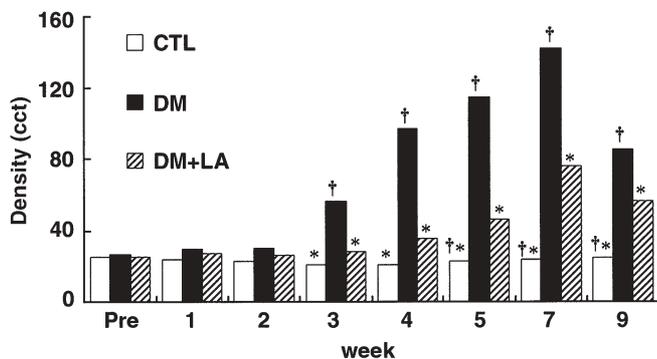


Figure 2. Changes in light scattering intensity at a depth of 0.08 mm. Light-scattering intensity in the DM group increased rapidly with time. Although the DM+LA group also showed an increase in light-scattering intensity, the change was much smaller than in the DM group. *cct*, computer compatible tape; □ *CTL*, control; ■ *DM*, diabetes mellitus; ▨ *DM+LA*, DM with α -lipoic acid group. * $P < 0.05$ versus DM. [†] $P < 0.05$ versus DM+LA.

Discussion

Although LA has been reported to prevent cataract development in rats treated with L-buthionine sulfoximine⁴ and in a sand rat model of acute type 2 diabetes,⁹ it has not been reported whether LA can prevent or delay cataract progression. The present study investigated the preventive effect

of LA on the development and progression of cataract in streptozotocin-induced diabetic rats during a 10-month period. Light-scattering measurements indicated that dietary LA is effective in delaying not only cataract development but also its progression. LA may be able to do this by preventing protein glycation¹² and reducing oxidative stress,^{1,3,7} two of the three putative mechanisms of diabetic cataractogenesis;³ LA is ineffective in reducing polyol pathway activity,^{3,14} the third mechanism.

Swamy-Mruthinti et al.¹⁵ measured progressive changes in lens opacification by Scheimpflug densimetric analysis and correlated them with protein glycation in the STZ-induced diabetic rat, and reported that crystallin glycation, rather than plasma glucose level, was strongly correlated with lens opacification. Suzuki et al.¹¹ also showed that LA significantly prevented BSA glycation and modification. Although we have not measured glycation in lens crystallin, it is possible that glycation in the DM+LA group may have been prevented by LA.

In the present study, LA showed an inhibitory effect on elevation of blood glucose at 80 and 111 days (Table 1). However, our results agree with those of previous studies.^{9,16} In some other studies,^{9,14,17–20} however, LA did not significantly lower blood glucose in diabetic conditions. This discrepancy may be due to (1) differences in the dosing and delivery of LA, (2) differences in animal models, or (3) differences in blood glucose sampling times. In fact, our results showed significantly lower blood glucose in DM+LA rats than in untreated DM rats only at 80 and 111 days. Kha-maisi et al.¹⁶ suggested that short-term administration of LA at high dosage to fasting normal or STZ-induced diabetic rats caused inhibition of systemic glucose production. The hypoglycemic effect of LA observed in our study may result from the above-mentioned mechanisms, and this effect may have partially contributed to ameliorating diabetic conditions. However, even when blood glucose levels were significantly lower in the DM+LA group than in the DM group, in agreement with a previous study,⁹ the levels were still far above the normoglycemic range. Therefore, the LA inhibitory effect on blood glucose elevation does not appear to be the main mechanism of prevention of diabetic cataract formation.

Acknowledgment. This study was supported by Sankyo Company Ltd., Tokyo, Japan.

References

1. Packer L, Witt EH, Tritschler HJ. α -Lipoic acid as a biological antioxidant. *Free Radic Biol Med* 1995;19:227–250.
2. Packer L, Kraemer K, Rimbach G. Molecular aspects of lipoic acid in the prevention of diabetes complications. *Nutrition* 2001;17:888–895.
3. Bron AJ, Brown NA, Harding JJ, Ganea E. The lens and cataract in diabetes. *Int Ophthalmol Clin* 1998;38:37–67.
4. Maitra I, Serbinova E, Trischler H, Packer L. α -Lipoic acid prevents buthionine sulfoximine-induced cataract formation in newborn rats. *Free Radic Biol Med* 1995;18:823–829.

LIPOIC ACID EFFICACY AGAINST RAT DIABETIC CATARACT

5. Maitra I, Serbinova E, Tritschler HJ, Packer L. Stereospecific effects of R-lipoic acid on buthionine sulfoximine-induced cataract formation in newborn rats. *Biochem Biophys Res Commun* 1996; 221:422–429.
6. Packer L. Antioxidant properties of lipoic acid and its therapeutic effects in prevention of diabetes complications and cataracts. *Ann N Y Acad Sci* 1994;738:257–264.
7. Kilic F, Handelman GJ, Traber K, et al. Modelling cortical cataractogenesis XX. In vitro effect of α -lipoic acid on glutathione concentrations in lens in model diabetic cataractogenesis. *Biochem Mol Biol Int* 1998;46:585–595.
8. Kilic F, Handelman GJ, Serbinova E, Packer L, Trevithick JR. Modelling cortical cataractogenesis 17: in vitro effect of α -lipoic acid on glucose-induced lens membrane damage, a model of diabetic cataractogenesis. *Biochem Mol Biol Int* 1995;37:361–370.
9. Borenshtein D, Ofri R, Werman M, et al. Cataract development in diabetic sand rats treated with α -lipoic acid and its gamma-linolenic acid conjugate. *Diabetes/Metab Res Rev* 2001;17:44–50.
10. Kilic F, Trevithick JR. Modelling cortical cataractogenesis. 16. Leakage of lactate dehydrogenase: a new method for following cataract development in cultured lenses. *Biochem Mol Biol Int* 1995;35:1143–1152.
11. Suzuki YJ, Tsuchiya M, Packer L. Lipoate prevents glucose-induced protein modifications. *Free Radic Res Commun* 1992;17: 211–217.
12. Ou P, Nourooz-Zadeh J, Tritschler HJ, Wolff S. Activation of aldose reductase in rat lens and metal-ion chelation by aldose reductase inhibitors and lipoic acid. *Free Radic Res* 1996;25:337–346.
13. Kojima M, Sasaki K. Application of a new Scheimpflug camera (EAS-1000) to animal cataract models. *Ophthalmic Res* 1992;24 Suppl 1:3–9.
14. Obrosova I, Cao X, Greene DA, Stevens MJ. Diabetes-induced changes in lens antioxidant status, glucose utilization and energy metabolism: effect of DL- α -lipoic acid. *Diabetologia* 1998;41:1442–1450.
15. Swamy-Mruthinti S, Green K, Abraham EC. Scheimpflug densitometric analysis of cataracts in diabetic rats: correlation with glycation. *Ophthalmic Res* 1996;28:230–236.
16. Khamaisi M, Rudich A, Potashnik R, et al. Lipoic acid acutely induces hypoglycemia in fasting nondiabetic and diabetic rats. *Metabolism* 1999;48:504–510.
17. Coppey LJ, Gellett JS, Davidson EP, Dunlap JA, Lund DD, Yorek MA. Effect of antioxidant treatment of streptozotocin-induced diabetic rats on endoneurial blood flow, motor nerve conduction velocity, and vascular reactivity of epineurial arterioles of the sciatic nerve. *Diabetes* 2001;50:1927–1937.
18. Cameron NE, Cotter MA, Horrobin DH, Tritschler HJ. Effects of α -lipoic acid on neurovascular function in diabetic rats: interaction with essential fatty acids. *Diabetologia* 1998;41:390–399.
19. Hounsom L, Horrobin DF, Tritschler H, Corder R, Tomlinson DR. A lipoic acid-gamma linolenic acid conjugate is effective against multiple indices of experimental diabetic neuropathy. *Diabetologia* 1998;41:839–843.
20. Nagamatsu M, Nickander KK, Schmelzer JD, et al. Lipoic acid improves nerve blood flow, reduces oxidative stress, and improves distal nerve conduction in experimental diabetic neuropathy. *Diabetes Care* 1995;18:1160–1167.